

# TwistAmp® exo RT Quick Guide

Part Number: TAEXORT02Guide | Revision A

## Basic Information

### RPA

- 1) Primers must be 30-35 bases
- 2) Works best at constant temperature (40°C)
- 3) Amplicons of 80-400bp are preferred
- 4) TwistAmp® exo Probe required - see overleaf

### PCR

- 1) Primers typically 18-25 bases
- 2) Thermal cycling required
- 3) Amplicons of 50bp upwards are typical/optimal

### Set-up (*single-plex*)<sup>1</sup>

- 1) Prepare reaction mix in 1.5ml tube:

Primer A (10µM)	2.1 µl
Primer B (10µM)	2.1 µl
TwistAmp® exo Probe (10µM)	0.6 µl
Rehydration Buffer	29.5 µl
Template,	
RNase Inhibitor and dH <sub>2</sub> O	13.2 µl
(Total Volume	47.5 µl)

Vortex and spin briefly
- 2) Add reaction mix to freeze-dried reaction. Pipette to mix.
- 3) Add 2.5 µl of 280mM MgAc (supplied) and mix well to start reaction.

**WARNING: RPA REACTIONS START AT ROOM TEMPERATURE AS SOON AS MAGNESIUM IS ADDED.**

- 4) Place strip in Twista® and start run: 40°C, 20 minutes.
- 5) Remove strip after 5-7 minutes, invert vigorously 8-10 times to mix & spin briefly, replace in Twista®.

**WARNING: IF TUBES ARE OPENED AFTER AMPLIFICATION THERE IS A GREAT RISK OF CONTAMINATION OF WORK SURFACES WITH AMPLICON! ENSURE THAT APPROPRIATE AVOIDANCE MEASURES ARE TAKEN!**  
**WARNING: SWITCH OFF HEATED LIDS BEFORE STARTING REACTIONS!**

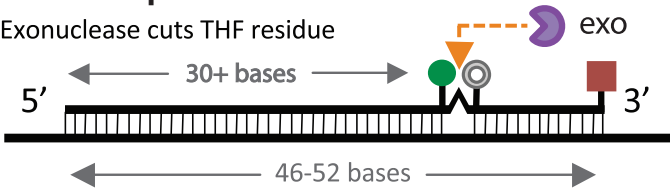
- 1 See manual for multiplexing

# RPA uses TwistDx's proprietary probe systems

RPA does NOT use PCR probe systems

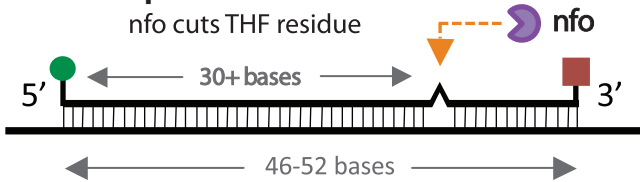
## TwistAmp® exo Probe

Exonuclease cuts THF residue



## TwistAmp® LF Probe

nfo cuts THF residue



● Fluorophore

● Nuclease

▲ THF residue

◎ Quencher

■ 3' block

*refer to manual for details of probe design*